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# Storage and recall of environmental signals in a plant: modelling by use of a differential (continuous) formulation

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## Abstract

The breaking of the symmetry of bud growth in *Bidens* seedlings involves a sort of plant 'memory'. An asymmetrical stimulus (e.g., the pricking of one of the seedling cotyledons) stores a 'symmetry-breaking' signal within the plants (function STO). Depending on other stimuli received by the seedlings, the stored signal may remain silent or be recalled (RCL function) and take effect in the seedling morphogenesis (asymmetry of the growth of the cotyledonary buds, with a statistical advantage to the bud at the axil of the non-stimulated cotyledon). We show that this memory mechanism can be interpreted by a model taking into account a genetic control exerted on a non-linear enzymatic system that is able to choose trajectories going to different attractors, depending on the stimulation intensity. **To cite this article: J. Demongeot et al., C. R. Biologies ●●● (●●●●).**

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## Résumé

**Mémorisation et rappel de signaux exogènes dans une plante : modélisation à l'aide d'une formulation (continue) différentielle.** La rupture de la symétrie de croissance des bourgeons de plantules de *Bidens* met en jeu une sorte de « mémoire » végétale. Un stimulus asymétrique (par exemple, l'application de quelques piqûres d'épingle à l'un des deux cotylédons de chaque plantule) induit le stockage (fonction STO) d'un message de « rupture de symétrie » dans les plantules. Selon que les plantules reçoivent ou non d'autres stimuli, le message stocké peut rester inexprimé ou être rappelé (fonction RCL) et s'exprimer dans la morphogenèse des plantules (asymétrie de la croissance des bourgeons cotylédonaire, avec un avantage statistique pour le bourgeon à l'aisselle du cotylédon non stimulé). Nous montrons que ce mécanisme de mémoire peut être interprété à l'aide d'un modèle prenant en compte le contrôle génétique exercé sur un système enzymatique non linéaire, capable de choisir des trajectoires conduisant à différents attracteurs, en fonction de l'intensité du stimulus. **Pour citer cet article : J. Demongeot et al., C. R. Biologies ●●● (●●●●).**

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**Keywords:** Differential equations; Memory; Evocation; Bud growth; Apical dominance; *Bidens pilosa* L.; Plants

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## 1. Introduction

In a previous paper [1], we have modelled the storage, recall and final effect of abiotic signals in seedlings of *Bidens pilosa* L. by use of an asynchronous, logical (discrete) formulation [2]. This was especially useful to determine toward which direction the system would tend to move and its steady-state conditions of functioning. The present paper represents the differential (continuous) approach of the same problem. The aim is to describe the system evolution by introducing a continuous term coming from non-linear enzyme kinetics into the equations of the dynamics of the seedling growth.

## 2. Summarizing the experimental data

See, e.g., Desbiez et al. [3–5] for original experimental data on the *Bidens* system and Thellier et al. [6] for a review. Briefly, the *Bidens* seedlings exhibit a bilateral symmetry with their two opposite cotyledons (cotyledon A on the left and cotyledon B on the right when all the seedlings under study are oriented parallel to one another), their actively growing apex (or terminal bud) and their two cotyledonary buds (i.e. the buds at the axil of the cotyledons). As long as the apex is present, it inhibits the growth of the ‘cotyledonary buds’ (apical dominance); however, after apex removal (seedling ‘decapitation’), the cotyledonary buds can start to grow. Stimulating (for instance, pricking) one of the seedling cotyledons (e.g., cotyledon A), i.e. delivering an asymmetrical stimulus to each seedling, tends to alter the symmetry of the growth of the ‘cotyledonary buds’. More precisely, the stimulation of one of the seedling cotyledons gives a statistical advantage to the axillary bud of the opposite cotyledon (the ‘distal’ bud) relative to the bud at the axil of the stimulated cotyledon (the ‘proximal’ bud). In a population of seedlings, this relative advantage of one bud upon the other can be measured by the use of a parameter,  $g$ , which corresponds to the difference (normalised to the total number of plants) between the number of plants where bud B starts growing before bud A and the number of plants where bud A starts growing before bud B. For instance,  $g = 1$  corresponds to the extreme asymmetrical response when bud B is the first to start to grow in all plants,  $g = -1$  to the other extreme asymmetrical response when bud A is the first to start to grow in all plants and  $g = 0$  to the symmetrical response in which there is an equal number of plants where it is bud A or bud B that is the first to start to grow.

When cotyledon stimulation is carried out on non-decapitated seedlings, this has no externally apparent

effect on plant morphogenesis; however, if the apical bud is finally removed, the cotyledonary buds start to grow with the same  $g$ -values as when cotyledon stimulation is carried out simultaneously to seedling decapitation. A ‘symmetry-breaking signal’ (initiated by cotyledon stimulation) thus is stored (‘STO’ function) within the seedlings, without taking effect, during the time lapse between cotyledon stimulation and seedling decapitation. From many experiments, it was inferred that this signal storage was an irreversible and an all-or-nothing process. Now, depending on the conditions of seedling decapitation and on the possible application of a variety of other treatments, the  $g$ -values appeared to be sometimes close to zero and sometimes significantly different from zero, although all the seedlings had been subjected to the same asymmetrical pricking treatment. This means that, apart from their STO function, the seedlings possess a recall (RCL) function that can be reversibly switched ‘on’ or ‘off’ and that the stored signal can take effect (differential growth of the cotyledonary buds) only when the RCL function is ‘on’. Typical experimental data taken from the first two Tables in Thellier et al. [6] are grouped in Table 1. By successively switching the recall function, ‘off’, then ‘on’, then ‘off’ again, etc., the observed  $g$ -values were close to zero, significantly above zero, close to zero again, etc. This shows that the stored symmetry-breaking signal can be repeatedly solicited, which is strikingly similar to the storage and evocation functions existing in animal and human brain [7,8].

## 3. The differential modelling of the *Bidens* system

### 3.1. Statement of the problem

The interpretation given below is based upon the assumption that the growth of any of the three buds of each seedling (the apex and the two cotyledonary buds) tends to further favour its own growth and to inhibit the growth of the other two buds, as a consequence of the synthesis by a growing bud of appropriate ‘growth metabolites’ [9].

### 3.2. The differential symbolism

In the present differential formulation, we are going to use specific symbols, which may be different from those used in the previous asynchronous, logical formulation [1]. In particular, here we are using the unique symbol A (respectively B) in order to represent, both, the left (respectively right) side of the seedlings (cotyledons and cotyledonary buds) and the size (i.e. also the

Table 1

Experimental results relative to bud growth asymmetry, following various sorts of pricking treatments, or combinations of pricking treatments, and plant decapitation [6]

Ref. No.	Pricking treatment	Plant decapitation at	$g$	Bud growth asymmetry
1	non-pricked control	onset of daylight	0.02	bud A $\approx$ bud B
2	4A	onset of daylight	$0.35 \pm 0.2$	bud B > bud A
3	4A	midday	$0.08 \pm 0.15$	bud A $\approx$ bud B
4	1A	onset of daylight	0.01	bud A $\approx$ bud B
5	4B	onset of daylight	-0.35	bud A > bud B
6	1A(1h)4B	onset of daylight	0.39	bud B > bud A
7	2A	onset of daylight	0.06	bud A $\approx$ bud B
8	2A(1h)2A/2B	onset of daylight	0.32	bud B > bud A
9	2A(1h)2A/2B(3h)2A/2B	onset of daylight	0.05	bud A $\approx$ bud B
10	2A(1h)2A/2B(3h)2A/2B(5h)2A/2B	onset of daylight	0.34	bud B > bud A

Meaning of symbols:  $x_A$ ,  $x$  pricks to cotyledon A;  $x_A/x_B$ ,  $x$  pricks simultaneously to both cotyledons A and B;  $x_A(yh)z_B$ ,  $x$  pricks to cotyledon A followed,  $y$  hours later, by  $z$  pricks to cotyledon B;  $g$  defined in text; bud A  $\approx$  bud B, no significant asymmetry; bud B > bud A, asymmetry in favour of bud B; bud A > bud B, asymmetry in favour of bud A. The experiments were carried out under the so-called 'standard' conditions [6].

## Mnesic 'opernet'

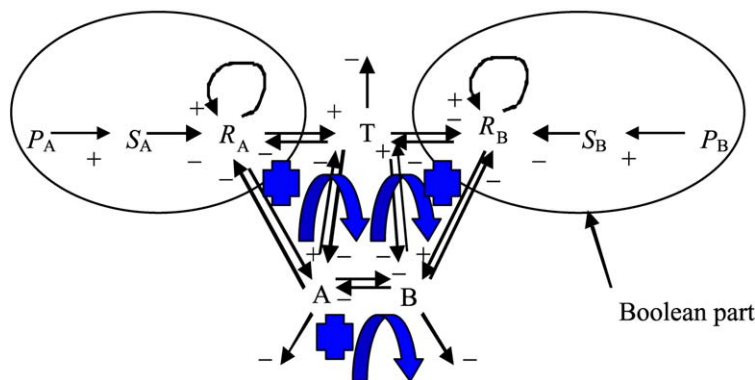


Fig. 1. The mnesic 'opernet' summarizing the regulation of the plant growth at two levels, the genetic control (Boolean part) and the epigenetic metabolism (differential part). The arrows + (respectively -) correspond to an activation (respectively an inhibition) and the blue arrows correspond to positive regulatory loops. (For interpretation of the references in colour in this figure legend, the reader is referred to the web version of this article.)

content in growth metabolites) of the cotyledonary buds. We term 'mnesic opernet' (Fig. 1) the system obtained by combining the genetic (*operon*) and epigenetic parts (metabolic *net*) of the system ruling the growth of the cotyledonary buds [6,8]. In Fig. 1, the genetic part of the mnesic opernet has been modelled using a Boolean system [1,8]: the variable  $P_A$  (respectively  $P_B$ ), which represents the pricking treatment (or any other asymmetrical stress action having an effect similar to that of pricking) on side A (respectively B) of the plant, is equal to 1 or 0, according to whether the asymmetrical treatment has been applied or not. The variable  $S_A$  (respectively  $S_B$ ) represents the discrete part of the opernet. The genetic control system is symbolized in Fig. 1 by  $S_A$  (respectively  $S_B$ ) on the side A (respectively B); moreover, we assume that  $S$ , as activated by the stimulus  $P$ , inhibits the expression of a morpho-

genetic cotyledonary material,  $R$ , which is responsible for the growth of both the apex and the cotyledonary bud A (respectively B). More precisely,  $R_A$  (respectively  $R_B$ ) represents the concentration of  $R$  on side A (respectively B) and we assume that this concentration  $R_A$  (respectively  $R_B$ ) is continuous and that the rate of  $R$  modification,  $dR_A/dt$  (respectively  $dR_B/dt$ ) is ruled by a differential system containing a 3-switch between the continuous variables  $T$  (size of the terminal bud),  $A$  and  $B$  (sizes of the cotyledonary buds). It may be recalled that an  $n$ -switch is a fully connected system of  $n$  elements with only negative interactions (inhibitions) between one another, except for possible positive autocatalytic terms. If there are no positive autocatalytic terms, and when  $n$  is an odd number, the  $n$ -switch system contains both positive and negative circuits; for instance, a 3-switch system contains two negative circuits

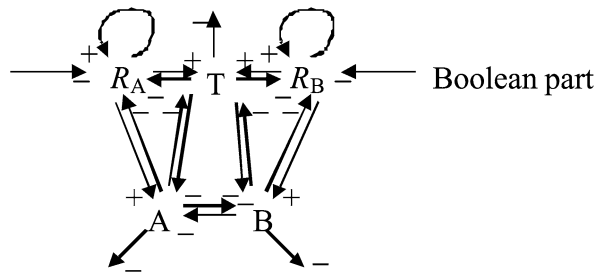


Fig. 2. Interaction graph  $G$  of the epigenetic part representing the metabolic core of the growth system. The central regulation triangle between  $T$ ,  $A$  and  $B$  is an inhibitory 3-switch.

(each made of three elements) and three positive circuits (each made of two elements). By contrast, all the circuits are positive when  $n$  is an even number. The study of the kinetics of such 3-switch systems [10] has shown that in most cases the asymptotic dynamical behaviour of growth consists of favouring the growth of one element of the  $n$ -switch and inhibiting the others.

Fig. 2 describes the interaction graph denoted  $G$ . The connectivity coefficient of this graph (i.e. the number of interaction arrows divided by the number of vertices)  $K(G) = 16/5$ , the mean inhibition weight (i.e. the proportion of inhibitory interactions, symbolised by negative arrows, vs. the total number of interactions)  $I(G) = 10/16$ , the number of positive circuits (i.e. the number of closed pathways with an even number of inhibitions)  $P(G) = 5$ . Moreover,  $G$  is a connected graph (i.e. there exists at least one pathway between each pair of vertices) that contains two *regulons* (i.e. two sub-graphs, each with only two vertices and three arrows, one of the arrows being a self-activation loop and the other arrows making a negative loop [11]).

### 3.3. The system of differential equations

The differential system governing the variables  $T$ ,  $A$ ,  $B$ ,  $R_A$  and  $R_B$  is written:

$$\begin{aligned} dR_A/dt &= (\sigma - kT - (4k/5)A - (k/5)B)R_A \\ &\quad - F(R_A)[T + (4/5)A + (1/5)B]/(T + A + B) \\ &\quad - (wP_A(t) + wP_B(t)/2) \\ dR_B/dt &= (\sigma - kT - (4k/5)B - (k/5)A)R_B \\ &\quad - F(R_B)[T + (1/5)A + (4/5)B]/(T + A + B) \\ &\quad - (wP_B(t_p) + wP_A(t_p)/2) \\ dT/dt &= (F(R_A) + F(R_B))T/(T + A + B) - \nu T \\ dA/dt &= F(R_A)(4/5)A/(T + A + B) \\ &\quad + F(R_B)(1/5)B/(T + A + B) - \nu A \end{aligned}$$

$$\begin{aligned} dB/dt &= F(R_B)(4/5)B/(T + A + B) \\ &\quad + F(R_A)(1/5)A/(T + A + B) - \nu B \end{aligned}$$

The first two equations correspond to the dynamics of  $R_A$  (respectively  $R_B$ ). The derivative at time  $t$ ,  $dR_A(t)/dt$  (respectively  $dR_B(t)/dt$ ) results from the combination of four terms:

- (i) an auto-catalytic term,  $\sigma R_A(t)$  (respectively  $\sigma R_B(t)$ ), in which  $\sigma$  is a kinetic constant;
- (ii) a negative term,  $-kT(t) - (4k/5)A(t) - (k/5)B(t)R_A(t)$  (respectively  $-kT(t) - (4k/5)B(t) - (k/5)A(t)R_B(t)$ ), expressing the mutual inhibitions of buds  $T$ ,  $A$  and  $B$  by one another, in which  $k$  is a kinetic constant. Note that this inhibition is supposed for the buds to be more important on the same side ( $4k/5$ ) than on the other side ( $k/5$ );
- (iii) a positive term, corresponding to the production of growth metabolites  $T(t)$ ,  $A(t)$  and  $B(t)$ ,  $F(R_A(t))[T(t) + (4/5)A(t) + (1/5)B(t)]/(T(t) + A(t) + B(t))$  (respectively  $F(R_B(t))[T(t) + (1/5)A(t) + (4/5)B(t)]/(T(t) + A(t) + B(t))$ ). In this term, it is assumed that  $R_A$  (respectively  $R_B$ ) consumption has a non-linear kinetics described by the function  $F$  and that it presents a Michaelian competitive inhibition by the bud both on its own side,  $A$  (respectively  $B$ ), and on the other side,  $B$  (respectively  $A$ ). We also assume that the  $V_{max}$  of the Michaelian inhibition is equal to 1 and that the affinity and competitive coefficients are equal to 1 for  $T$  and to  $4/5$  and  $1/5$  for  $A$  (respectively  $B$ ), depending on the side on which they exert their inhibition;
- (iv) instantaneous perturbations  $wP_A(t)$  from side  $A$  (respectively side  $B$ ) and  $wP_B(t)/2$  from the other side  $B$  (respectively  $A$ ), the value of  $P_A(t)$  (respectively  $P_B(t)$ ) being 1 if the pricking treatment is applied to cotyledon  $A$  (respectively cotyledon  $B$ ) at time  $t = t_p$ , and 0 anywhere else. The coefficient  $w$  quantifies the intensity of the perturbations, supposed to be equal to  $w$  on the same side and  $w/2$  on the opposite side.

Note that, in these first two equations, the coefficients  $4/5$  and  $1/5$ , and  $1$  and  $1/2$ , have been chosen arbitrarily, just to express that the effect of one bud upon itself is much larger than its effect upon the other bud. Given the robustness of the system, the results would not be changed significantly by choosing other parameter values (e.g.,  $5/6$  and  $1/6$ , and  $1$  and  $1/3$ ).

The last three equations correspond to the dynamics of the concentrations of growth metabolites in the

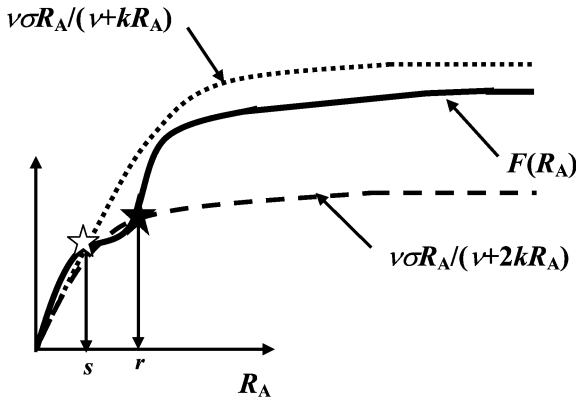


Fig. 3. The function  $F(R_A)$  presenting two successive inflection points (full curve), the function  $v\sigma R_A/(v+kR_A)$  (dotted curve) and the function  $v\sigma R_A/(v+2kR_A)$  (dashed curve). The black star corresponds to a stable steady state of abscissa  $R_A = r$  and the white star to an unstable steady state of abscissa  $R_A = s$ .

apex and the cotyledonary buds,  $dT(t)/dt$ ,  $dA(t)/dt$  and  $dB(t)/dt$ . These equations merely express the fact that the production of the growth metabolites depends on  $R_A(t)$  and  $R_B(t)$ , with a competitive inhibition by the other sources of growth as already described above, plus a linear degradation term, the kinetic constant of which is equal to  $\nu$ .

In order to be able to interpret a minima the experimental results given in Table 1, let us assume that  $F$  is a ‘dual-phasic’ function possessing two successive inflection points [12] (Fig. 3). Moreover, dual-phasic kinetic curves have also been shown to exist with natural or artificial enzymatic or transport systems [13–15].

Let us consider the stationary state equation of the differential system as described above. In the absence of any perturbation, it is written:

$$\begin{aligned} dR_A/dt = 0, \quad dR_B/dt = 0, \quad dT/dt = 0, \\ dA/dt = 0, \quad dB/dt = 0 \end{aligned}$$

If the value of the derivative of  $F$ ,  $F'(R_A) = dF(R_A)/dR_A$ , verifies  $F'(0) > \sigma$ ,  $F'(r) > \nu^2\sigma/(\nu + 2kr)^2$ ,  $F'(s) > \nu^2\sigma/(\nu + ks)^2$ , with  $F(r) = \nu\sigma r/(\nu + 2kr)$ ,  $F(s) = \nu\sigma s/(\nu + ks)$ , then the differential system as indicated above possesses, in the space  $(R_A, R_B, T, A, B)$ , among all its stationary states, (i) the stable steady state  $(0, 0, 0, 0, 0)$ , (ii) two unstable focuses  $(s, 0, \sigma s/(\nu + ks), 0, 0)$  and  $(0, s, \sigma s/(\nu + ks), 0, 0)$  (which we assume to be surrounded by the limit cycles denoted  $\alpha$  and  $\beta$ , respectively) and (iii) the stable steady state  $C$ , which is equal to  $(r, r, 2\sigma r/(\nu + kr), 0, 0)$  (Fig. 4).

In our reasoning below, we assume that the dynamics of the inhibitory 3-switch representing the interactions

between  $T$ ,  $A$  and  $B$  is already known [10]. Note that, if we consider an inhibitory  $n$ -switch between the entities  $A_i$ 's and if this  $n$ -switch verifies:

$$dA_i/dt = \left( \sum_{j=1, \dots, n} K_{ij} A_j \right) / \left( \sum_{j=1, \dots, n} A_j \right) - \nu A_i$$

then the stationary states  $A_i = K_{ii}/\nu$ ,  $A_k = 0$ , for  $k \neq i$  are stable. It is easy to check this property on the Jacobian matrix of the differential system. See Cinquin and Demongeot [10] for a complete study of the inhibitory  $n$ -switches. Briefly, such a system has a single non-zero real eigenvalue  $\lambda = -\nu < 0$  (all the other eigenvalues being equal to 0). This implies the stability, because of the negativity of all the second-order derivatives  $\partial^2(dA_k/dt)/\partial A_i^2$ , for  $k \neq i$ . In the following, we are going to assume that we start from a stable steady state  $(T(\infty) > 0, A(\infty) = B(\infty) = 0)$ ; when  $T$  will be given the value 0 by apex removal, then fluctuations will impose at time  $\tau$  the apparition of a non-zero value of either  $A(\tau)$  or  $B(\tau)$ , the probability of occurrence of both non-zero values being considered as negligible. Three cases may then be considered:

- (i) if  $R_A(\tau)$  and  $R_B(\tau)$  have approximately the same value, such as:

$$\begin{aligned} & F(R_A(\tau))(4/5)A(\tau)/(T(\tau) + A(\tau) + B(\tau)) \\ & + F(R_B(\tau))(1/5)B(\tau) \\ & / (T(\tau) + A(\tau) + B(\tau)) > \nu A(\tau) \\ & F(R_B(\tau))(4/5)B(\tau)/(T(\tau) + A(\tau) + B(\tau)) \\ & + F(R_A(\tau))(1/5)A(\tau) \\ & / (T(\tau) + A(\tau) + B(\tau)) > \nu B(\tau) \end{aligned}$$

then the steady state is  $A(\infty) > 0$  and  $B(\infty) = 0$ , if  $A(\tau) > 0$  and, inversely,  $B(\infty) > 0$  and  $A(\infty) = 0$ , if  $B(\tau) > 0$ . In this case, after seedling decapitation both cotyledonary buds,  $A$  and  $B$ , have equal chances to be the first to start to grow (thereby inhibiting the growth of the opposite bud) if both fluctuations have the same chance to occur;

- (ii) if  $R_A(\tau) \gg R_B(\tau)$  and  $A(\tau) > 0$ ,  $B(\tau) > 0$  or  $B(\tau) > 0$ ,  $A(\tau) > 0$ , such as:

$$\begin{aligned} & F(R_A(\tau))(4/5)A(\tau)/(T(\tau) + A(\tau) + B(\tau)) \\ & + F(R_B(\tau))(1/5)B(\tau) \\ & / (T(\tau) + A(\tau) + B(\tau)) > \nu A(\tau) \\ & F(R_B(\tau))(4/5)B(\tau)/(T(\tau) + A(\tau) + B(\tau)) \\ & + F(R_A(\tau))(1/5)A(\tau) \\ & / (T(\tau) + A(\tau) + B(\tau)) < \nu B(\tau) \end{aligned}$$

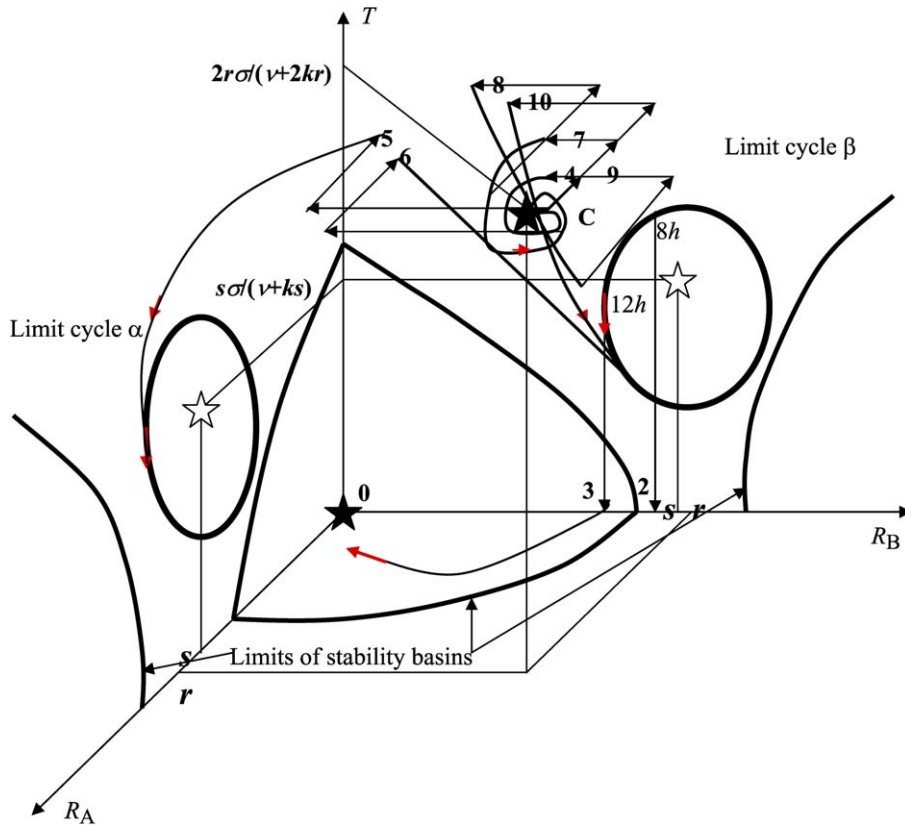


Fig. 4. Experimental perturbations in the state space  $(R_A, R_B, T)$ . See explanation in the text.

then the steady state is  $A(\infty) > 0, B(\infty) = 0$ , if  $A(\tau) > 0$  and no fluctuation  $B(\tau) > 0$  can be amplified. In this case, bud A takes precedence over bud B;

- (iii) the third case is identical to the second case, except that the roles of A and B have been inverted, and it is bud B that takes precedence over bud A.

#### 4. Application to the interpretation of the experimental data

When examining the experimental data represented in Table 1, the phenomenology of the growth response of the cotyledonary buds looks rather intricate: (i) the asymmetry of the response ( $g \neq 0$ ) depends not only on the intensity of the asymmetrical pricking treatment (1, 2 or 4 pricks), but also on the time of the day when plant decapitation was carried out, (ii) the combination of a pricking treatment 1A (which did not induce any bud growth asymmetry per se) with a treatment 4B (which induced an asymmetry in favour of bud A per se) induced an asymmetry in favour of bud B, and (iii) combining a treatment 2A with one or several symmetrical

treatments 2A/2B caused the bud growth response to be symmetrical (when there was an even number of treatments 2A/2B) or asymmetrical in favour of bud B (with an odd number of treatments 2A/2B). However, the theoretical model described in Fig. 4 gives a straightforward interpretation of the various aspects (see the corresponding reference numbers in Fig. 2 and in the first column of Table 1) of this apparently complicated phenomenology:

- (i) if the initial conditions (non-pricked controls) are within the basin of stability of the stable steady state C (i.e. close enough to C), then, in the absence of any perturbation, the whole trajectory lies within this basin and tends towards C (where  $R_A \approx R_B$ ) when time  $t$  increases to infinity. After seedling decapitation and in the absence of any previous pricking (non-pricked control), buds A and B have equal chances to be the first to start to grow, thereby inhibiting the growth of the opposite bud, with the consequence that  $g \approx 0$ ;
- (ii)–(iii) with seedlings subjected to a 4-pricks treatment on cotyledon A, there is a shift from the initial

conditions close to C (non-pricked controls) to the basin of the limit cycle  $\beta$ . That the final response depends on the time of the day when seedling decapitation was carried out ( $g > 0$  or  $g \approx 0$ , according to whether the seedlings were decapitated at the onset of daylight or in the middle of the day) is due to the fact that, after seedling decapitation,  $R_A$  and  $R_B$  lie within the basin of the limit cycle  $\beta$  in which  $R_B \gg R_A$  in the first case, while they are in the basin of the stable steady state 0 in the second case, in which  $R_B$  can be close to  $R_A$  and sufficiently large for obeying condition (1) at the end of Section 3. If the trajectory is slowly tending toward the stable steady state 0, haphazard fluctuations can cause either bud A or B to start to grow, with the consequence that  $g \approx 0$ ;

- (iv) if the system is initially close to C and if a 1-prick treatment is applied to cotyledon A (respectively B), then we assume that this perturbation results in an instantaneous decrease  $w$  of the intensity of  $R_A$  (respectively  $R_B$ ) and in a smaller decrease (e.g.,  $w/2$ ) of the intensity of  $R_B$  (respectively  $R_A$ ) at the time  $t_p$  when the pricking treatment is carried out. In such a case, the variables  $R_A$  and  $R_B$  remain in the basin of C and  $g \approx 0$ ;
- (v) when a 4-prick treatment is applied to cotyledon B, then the point representing the system leaves the basin of C with a decrease of, for instance,  $4w$  in  $R_A$  and  $2w$  in  $R_B$ , to go to the basin of the limit cycle  $\alpha$  in which  $R_A \gg R_B$ , hence  $g < 0$ ;
- (vi) when the seedlings are subjected to a 1-prick treatment on cotyledon A followed, 1 h later, by a 4-prick treatment on cotyledon B, then the system goes into the basin of the limit cycle  $\beta$ , with the consequence that  $R_B \gg R_A$  and  $g > 0$ ;
- (vii) when the seedlings are subjected to a 2-prick treatment on cotyledon A, then the system remains in the basin of C and  $g \approx 0$ ;
- (viii) when the seedlings are subjected to a 2-prick treatment on cotyledon A followed, 1 h later, by a symmetrical 2-prick treatment on both cotyledons A and B, then the system goes into the basin of the limit cycle  $\beta$ , again with the consequence that  $R_B \gg R_A$  and  $g > 0$ ;
- (ix) when the seedlings are subjected to a 2-prick treatment on cotyledon A followed, 1 h later, by a symmetrical 2-prick treatment on both cotyledons A and B, again followed, 3 h later, by a symmetrical 2-prick treatment on both cotyledons A and B, then the system returns into the basin of C with the consequence that  $g \approx 0$ . This is due to the special shape of the trajectory (8) that passes close

to the frontier of the basin of attractor C when going towards the limit cycle  $\beta$ ;

- (x) when a 2-prick treatment on cotyledon A is followed by three symmetrical 2-pricks treatments on both cotyledons A and B after 1 h, 3 h and 5 h, respectively, the system goes into the basin of C after the second symmetrical pricking treatment (same behaviour as in case (ix)), but it leaves the basin of C to go towards the limit cycle  $\beta$  after the third symmetrical treatment. In that case, we have again  $g > 0$ .

It is noteworthy that our above reasoning still holds if C is an attractor (e.g., a limit cycle). Because of its qualitative character, this reasoning is robust and depends only on the general features that have been chosen for function  $F$ : in particular, its dual-phasic non-linearity is absolutely requisite to be able to account for all the experimental observations.

## 5. Conclusions

By using the qualitative description of both the genetic (after a pricking treatment or other stresses) and epigenetic forces exerted on the opernet, all the simulated behaviours were shown to fit qualitatively the observed phenomenology as it is summarized in Table 1. In our present interpretation (i) the storage (STO) function corresponds to the existence of the limit cycles  $\alpha$  and  $\beta$  in the state space  $(R_A, R_B, T)$ , which can be considered as internal metabolic clocks of the growth function, while (ii) the recall (RCL) function depends on each particular pathway that a perturbation can impose to the point representing the system in its state space according to each particular condition of experiment. We can finally note, as already suggested in Tonnelier et al. [16], that exogenous instantaneous perturbations exerted on a non-linear system can both store and evoke information and therefore seem to be a necessary component of any memory system.

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